Using DNA Stable Isotope Probing to Identify MTBE and TBA Degraders In Situ

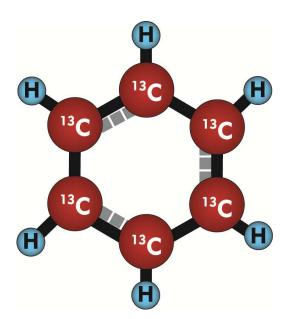
Katherine Clark and Dora Ogles *Microbial Insights, Inc.* Kerry Sublette *University of Tulsa* Kathleen Duncan *University of Oklahoma* Doug MacKay and Kate Scow *University of California at Davis*





Stable Isotope Compounds

- Specially produced "heavy" compounds which are composed of 99+% ¹³C
 - Natural compounds are 1.1% $^{\rm 13}{\rm C}$
 - Very similar chemical and biological characteristics as original compound
- Used as "tracers" to increase our understanding of contaminant fate



How do we do stable isotope probing?

- Obtain a ¹³C-labeled compound from a vendor
- Introduce the compound to the environment of interest and incubate
 - Soil
 - Groundwater
 - Microcosm
- Recover ^{13}C -labeled material and look for ^{13}C (usually involves GC-IRMS)
 - Residual compound
 - To assess biodegradation
 - Biomolecules of interest
 - DNA or RNA
 - Phospholipids
 - Proteins (¹³C or ¹⁵N)
 - Biodegradation products
 - Carbon dioxide or methane

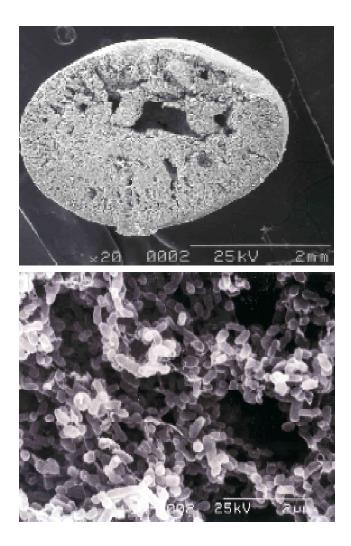
What do we learn from stable isotope probing?

- Finding ¹³C in biomass (usually membrane lipids) or biodegradation products indicates that biodegradation potential for the labeled compound exists in the sampled environment and that potential was expressed under *in situ* conditions (soil and groundwater)
- What organisms are involved in processing a ¹³C-labeled compound?
 - DNA
 - ¹³C- and ¹²C-DNA separated by ultracentrifugation
 - Sequencing of ¹³C-labeled 16SrDNA genes can identify organisms involved in processing the ¹³C-labeled compound
 - Differential labeling of a compound can even help identify microbes involved in processing different functional groups

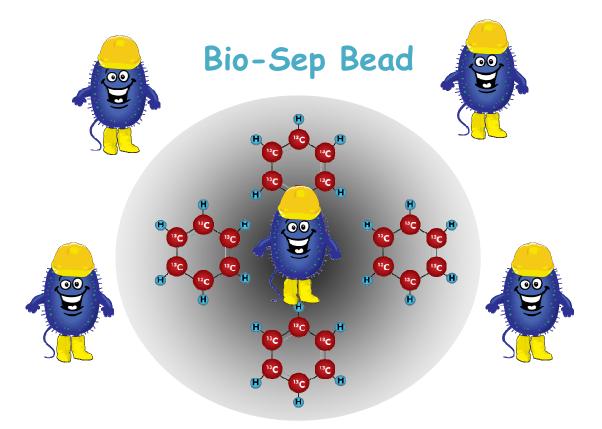
Anchoring the ¹³C-labeled compound

- Limiting dilution of the stable isotope during incubation in soil or groundwater greatly increases sensitivity of the method
 - Adsorption to activated carbon
 - Entrapment
- Bio-Sep[®] beads
 - 3-4 mm in diameter
 - 25% Nomex and 75% powered activated carbon
 - 600 m² of surface area
 - Heat sterilized
 - Colonized by actively growing microorganisms

Bio-Sep[®] beads and Bio-traps



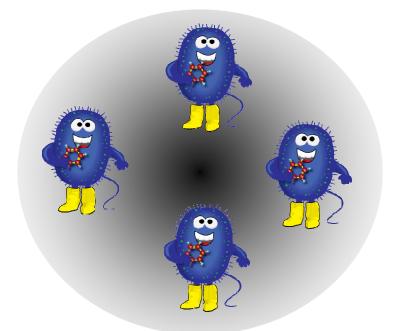




¹³C-labeled compounds sorbed to Bio-Sep[®] beads Bio-Trap colonized by indigenous microorganisms

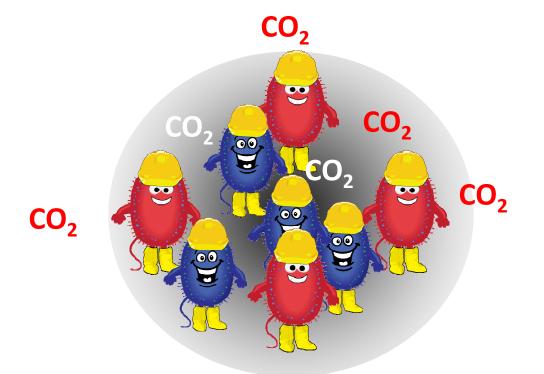
Microbes utilize target compound

Bio-Sep Bead



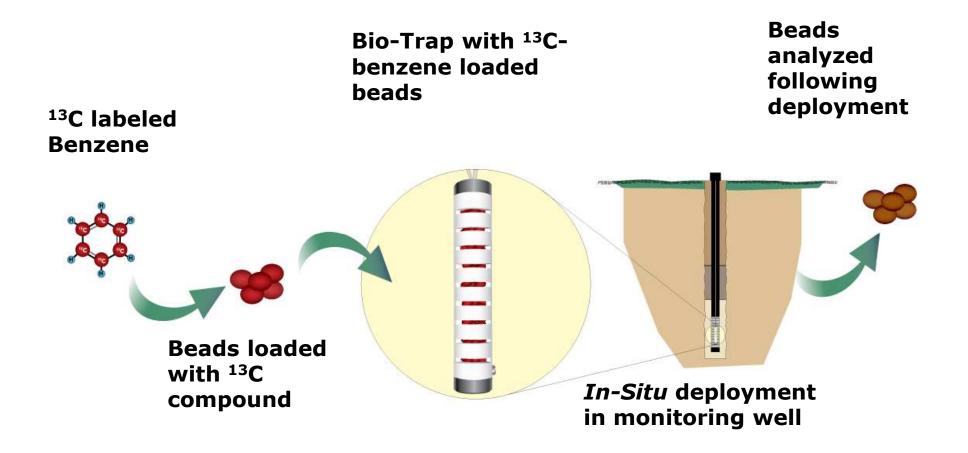
Some microbes that colonized the Bio-Sep[®] bead can utilize ¹³C labeled target compound.

^{13}C incorporation into biomass and CO_2

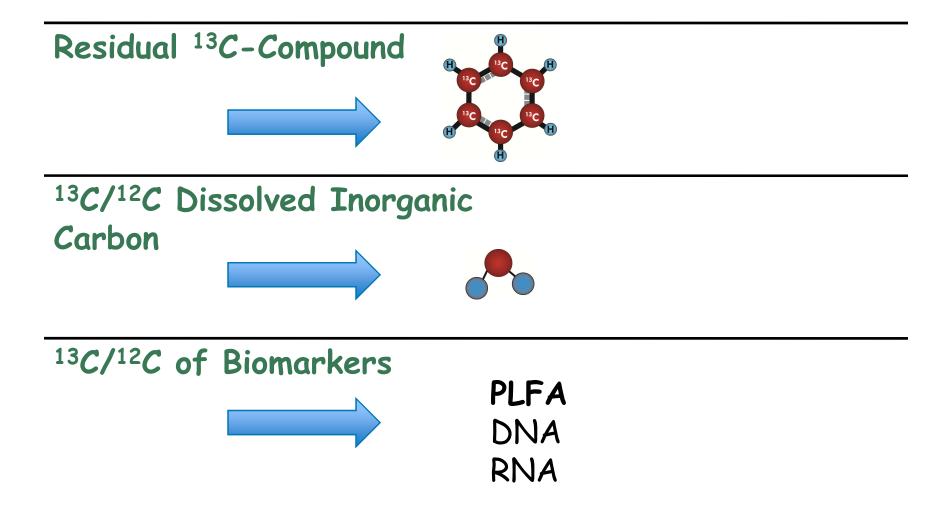


 $^{13}\mathrm{C}$ is incorporated into new cells growing in the beads and in CO_2

Overview of Bio-Trap SIP Approach

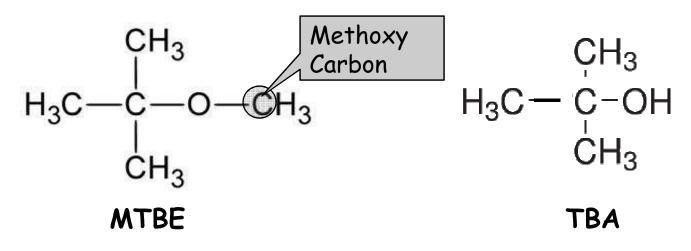


Bio-Trap SIP Analysis



Anaerobic MTBE and TBA Biodegradation

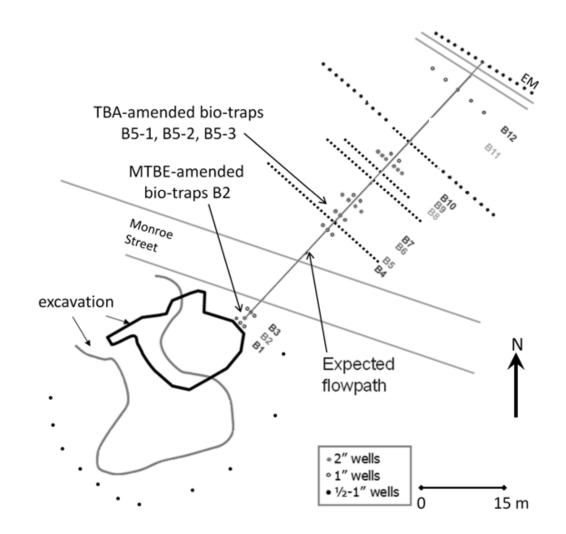
- No microorganisms capable of degrading MTBE or TBA anaerobically have been isolated
- O-demethylation thought to be first step in anaerobic MTBE degradation (acetogens)



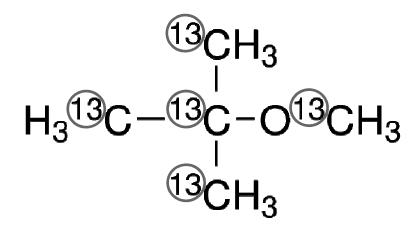
• TBA accumulation commonly observed

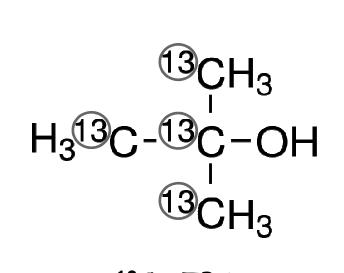
Vandenburg Air Force Base, Site 60

- 572 gal gasoline leak in 1994
- Source area excavation in 1995 and 2007
- Within 10 years BTEX had been degraded, but MTBE and TBA persisted
- Anaerobic, sulfatereducing aquifer

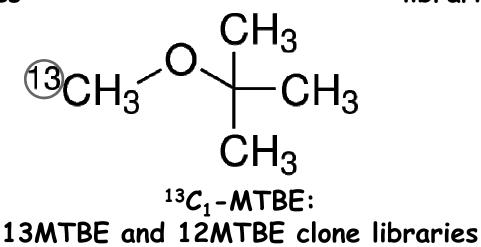


Contaminants used for DNA-SIP

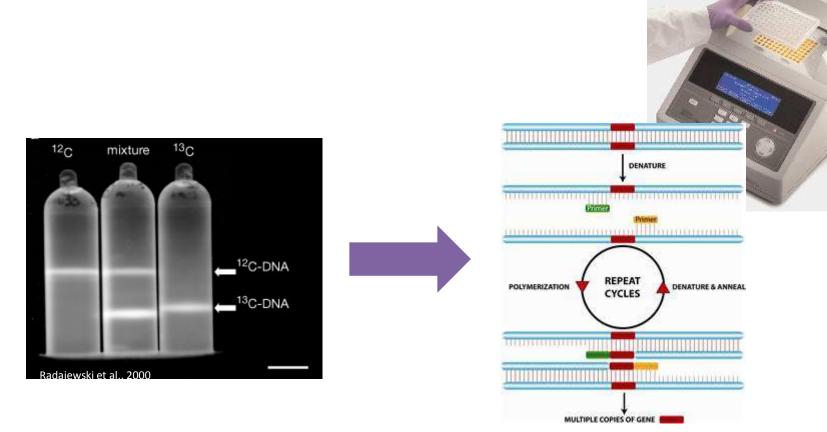




¹³C₅-MTBE: 13MALL and 12MALL clone libraries ¹³C₄-TBA: 13TBA and 12TBA clone libraries





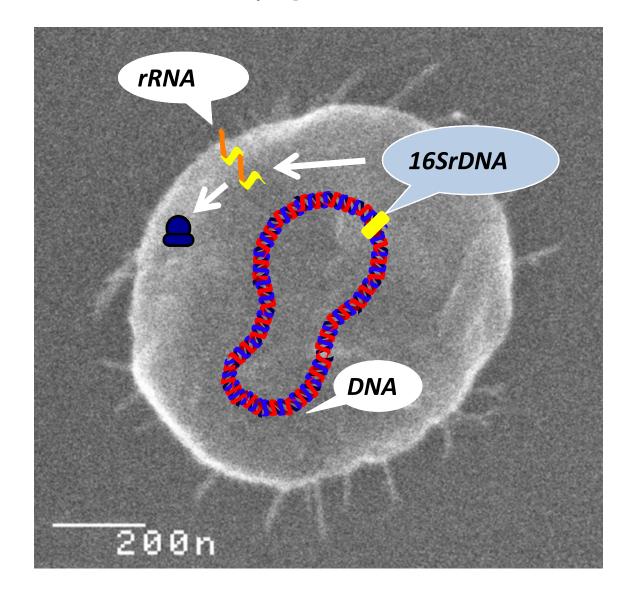


http://www.gbiosciences.com/EducationalUploads/Educational ProductsImages/mediumimages/PCR%20scheme.jpg

DNA was extracted from Bio-Sep® beads and ¹²C- and ¹³C-DNA are separated by CsCl density gradient centrifugation

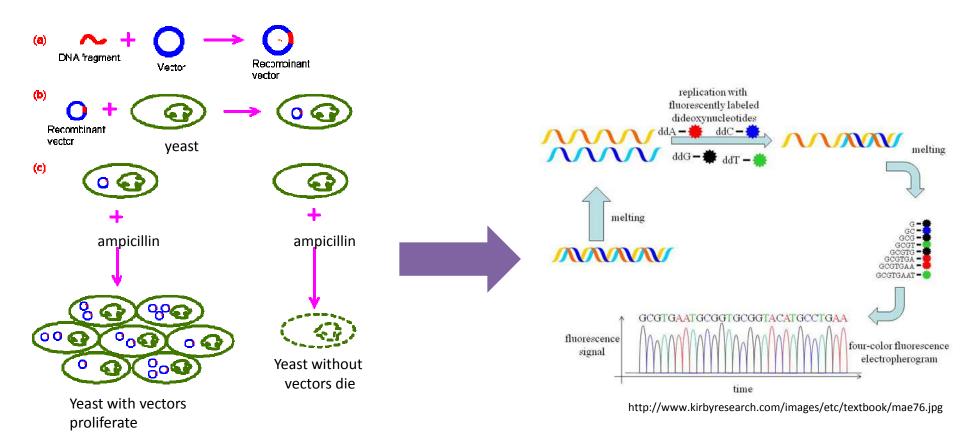
165 rRNA gene sequence of purified ¹²C- and ¹³C-DNA was amplified by PCR

16SrDNA is one of many genes in a microbe's DNA



The 16SrDNA gene is like an organisms business card





(d) isolation of recombinant DNA clones

http://www.web-books.com/MoBio/Free/images/Ch9A1.gif

Clone libraries were created from the PCR products and sequenced

Sequence Analysis Summary

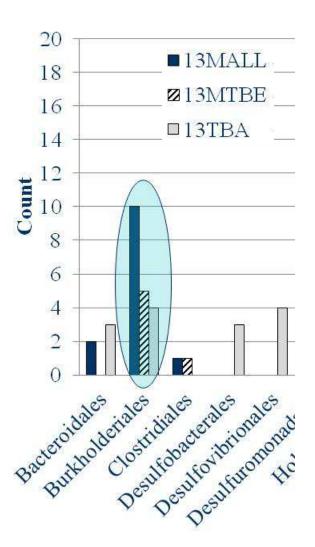
- 205 sequences were grouped into 60 OTUs
- Jaccard's Similarity Index indicated all clone libraries were dissimilar from each other

	#	#	%
	Sequences	OTUs*	Coverage**
13MTBE	41	7	93%
13MALL	42	20	69%
13TBA	42	23	69%
12MTBE	21	10	76%
12MALL	33	20	58%
12TBA	26	14	77%

*Operational taxonomic units based on 97% sequence similarity **Good's method

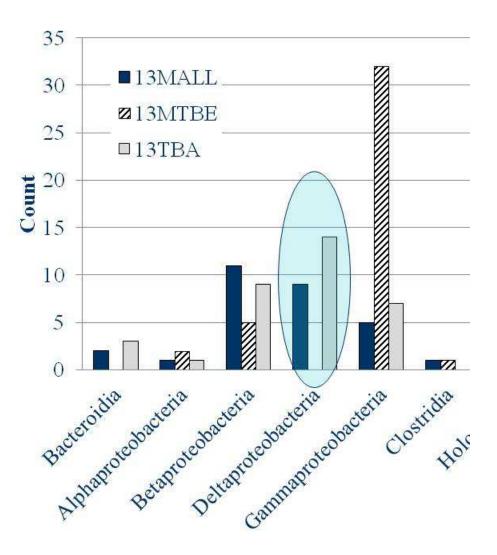
Sequences related to known MTBE and TBA degraders

- Many known MTBE and TBA degraders belong to the Burkholderiales order of the Betaproteobacteria, including PM1
- Members of *Sphingomonas* genus have been detected in anaerobic microcosms of MTBE-degrading cultures
- 1 *Sphingomonas* sequence in 13TBA and 2 in 13MTBE
- Likely to be primary degraders



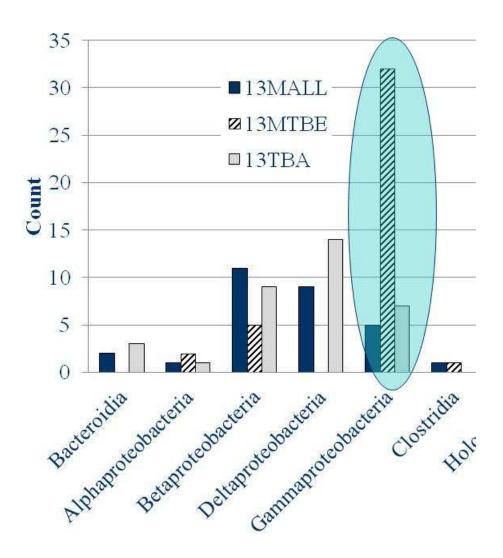
Deltaproteobacteria in 13MALL and 13TBA libraries

- Sulfate- and ironreducers likely involved in processing the *tert*-butyl group
- Desulfobulbus, Desulfovibrio, Desulfuromonas, and Geobacter belong to Deltaproteobacteria
- Geothrix also detected in 13TBA



Gammaproteobacteria predominate in 13MTBE library

- 19 sequences similar to
 Pseudomonas fluorescens
- May have crossfed on ¹³C-acetate
- *P. fluorescens* likely a secondary degrader



Conclusions

- Microorganisms degraded the ¹³C-labeled MTBE and TBA *in situ*, and ¹³C was incorporated into DNA
- OTUs belonging to order Burkholderiales and genus Sphingomonas were related to known MTBE and TBA degraders and most likely primary degraders
- Sulfate- and iron-reducers were likely involved in processing the *tert*-butyl group
- *P. fluorescens* detected in 13MTBE (only methoxy carbon labeled) was likely a secondary degrader, cross-feeding on metabolites, likely acetate

Questions?



Unit of measure

Amount of $~^{13}\text{C}$ relative to ^{12}C is expressed by the $\delta^{13}\text{C}$ notation

$$\delta^{13}C[\%_{0}] = \left(\frac{({}^{13}C/{}^{12}C)_{\text{Sample}}}{({}^{13}C/{}^{12}C)_{\text{Standard}}} - 1\right) \cdot 1000$$

The standard is a specific carbon-containing mineral from a specific location: Pee Dee Belimnite (PDB)

Units of
$$\delta^{13}C$$
 are °/₀₀ or "per mill"