# An *in situ* bioreactor for the treatment of groundwater impacted by petroleum hydrocarbons

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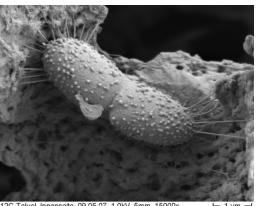
## Adapting Bio-Sep technology for remediation

- Bio-Sep beads have long been used a diagnostic tool to monitor in situ microbial activity
- Recently, trials have begun to test the effectiveness of a bioreactor utilizing Bio-Sep beads to bolster indigenous microbial populations and enhance bioremediation



## What is a Bio-Trap?

- Passive sampling tool for microorganisms
  - Simplest form consists of Bio-Sep beads in a housing that allows contact between the beads and groundwater
- Collects only environmentally competent, active microbes
  - Organisms must grow and reproduce within the beads to be detected
- Integrative sample (vs. snapshot)
  - Typical incubation 30 45 days
- Analyzed using molecular biological tools and stable isotope analysis



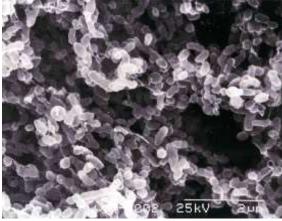
C-Toluol, Innenseite, 09.05.07, 1,0kV, 5mm, 150

⊢ 1 µm -

## How do Bio-Traps work? Bio-Sep beads

- 3-4 mm in diameter
- 25 % Nomex, 75% PAC
- 74% porosity
- 600 m<sup>2</sup> of surface area/g
- Surrounded by ultrafiltrationlike membrane with 1-10 micron holes
- Autoclavable
- Cleaned of fossil biomarkers by heating to 270 °C
- Biofilms form rapidly on Bio-Sep beads

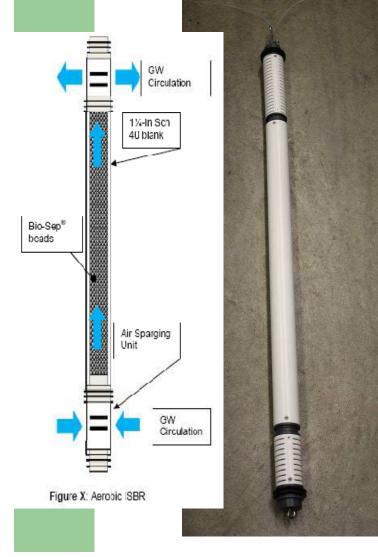




### **Characteristics of Bio-Sep beads useful for treatment applications**

- Provide favorable conditions for growth and reproduction
- Concentrate nutrients and hydrocarbons
  - Increasing local concentrations above threshold levels
  - Reducing concentrations of inhibitory aromatic hydrocarbons in the aqueous phase
- High surface area
- Rapid formation of biofilms
- Release new microbes into the aquifer once carrying capacity is reached

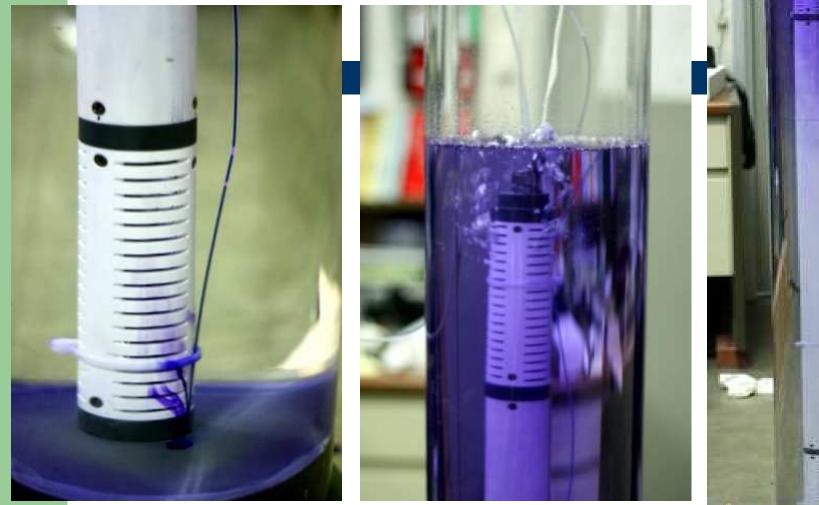
### Aerobic bioreactor design



- Fits in standard 2" well
- Packed bed bioreactor containing Bio-Sep beads open for fluid flow at top and bottom
- Air sparging into bottom of packed bed creates air lift for circulation of groundwater
- Air sparging and nutrient/sampling lines connected to surface equipment



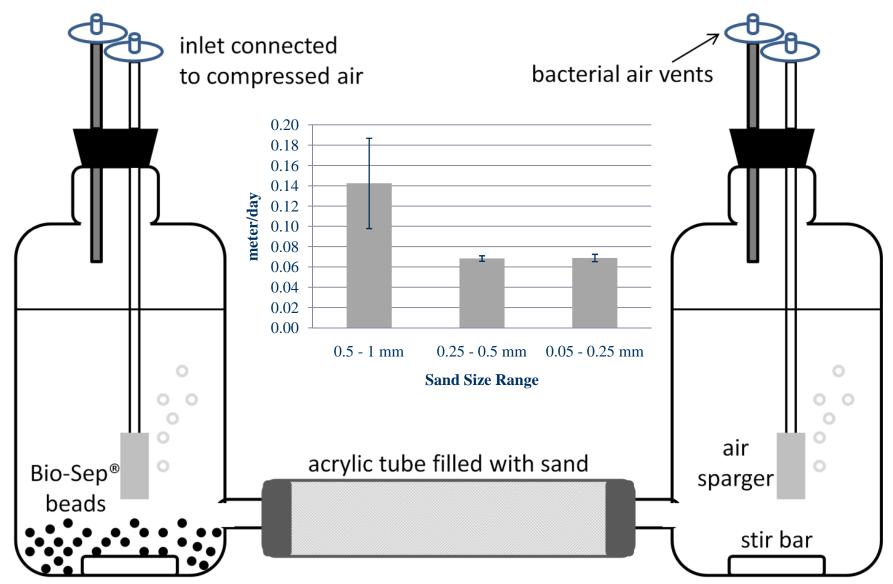
#### Aerobic bioreactor flow pattern





#### **Microbial release and transport**

outlet

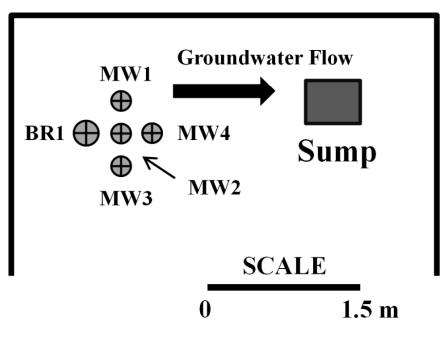


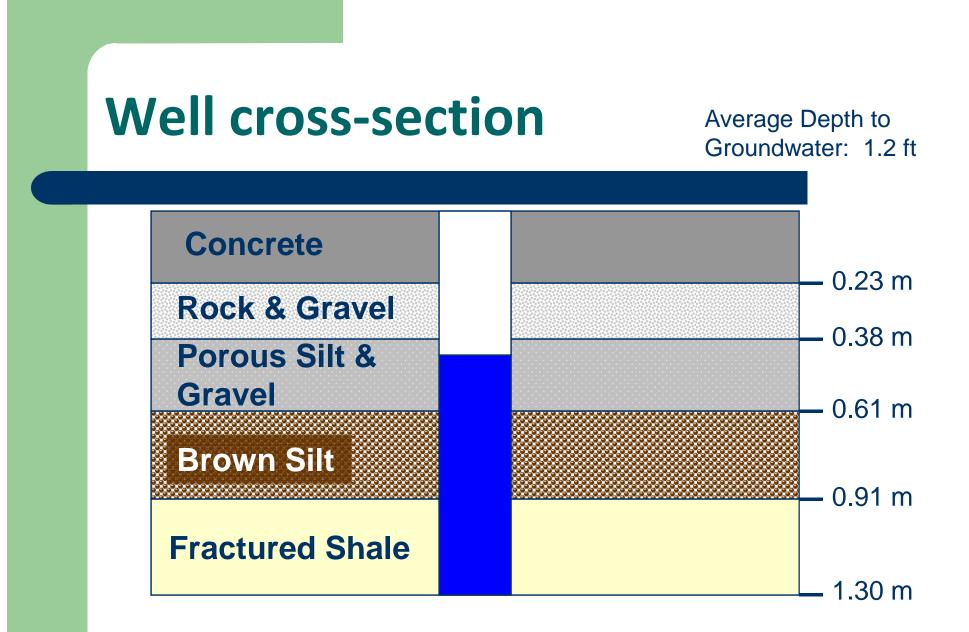
## **Operation of an aerobic bioreactor at hydrocarbon sites**

- Bio-Sep beads, nutrient addition, and air sparging encourage microbial growth and reproduction
- Contaminated groundwater is treated as it moves through the column of Bio-Sep beads
- Water exiting the reactor carries hydrocarbondegrading microbes into the aquifer

## Residential site impacted with heating oil

- Fuel oil release impacting soil and groundwater beneath a private residence
- Low, but persistent [BTEX]
- The bioreactor well and four monitoring wells were drilled in the basement
- Wells spaced 1 ft apart
- Fractured shale aquifer







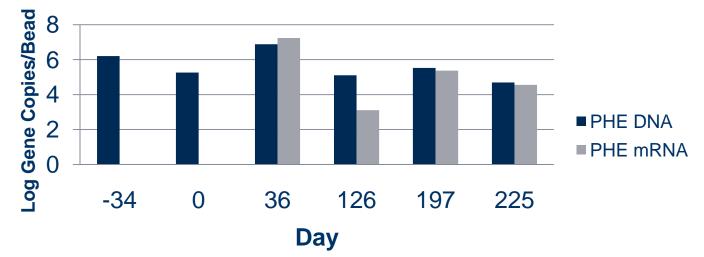
## **Bio-Trap Sampling**

- Bio-traps were deployed monthly in each of the 5 wells
- DNA and mRNA analyses
  - total eubacteria
  - functional genes associated with aromatic hydrocarbon degradation
    - Phenol Hydroxylase (PHE)
    - Naphthalene Dioxygenase (NAH)

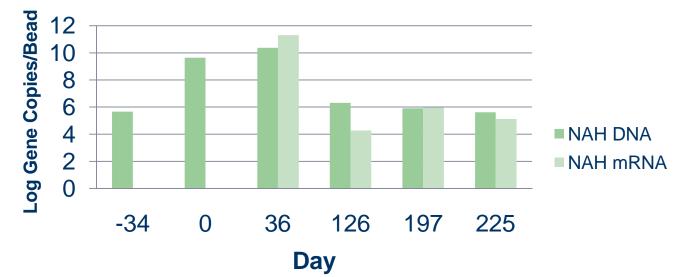
## **Bioreactor Timeline**

- Days -67 through -34: Air sparging only
- Days -34 through 0: Air sparging and nutrient delivery
- Day 0 onward: Complete bioreactor system operational

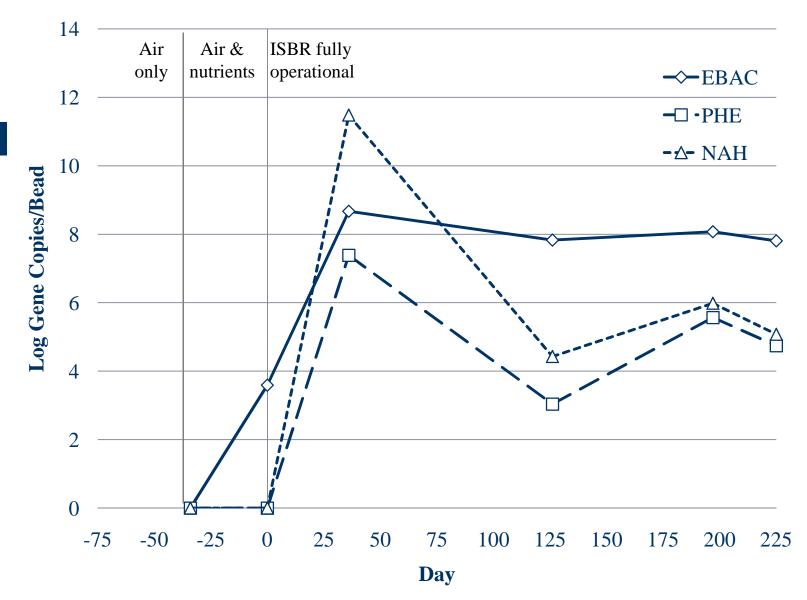
#### Comparison of PHE DNA and mRNA in BR1



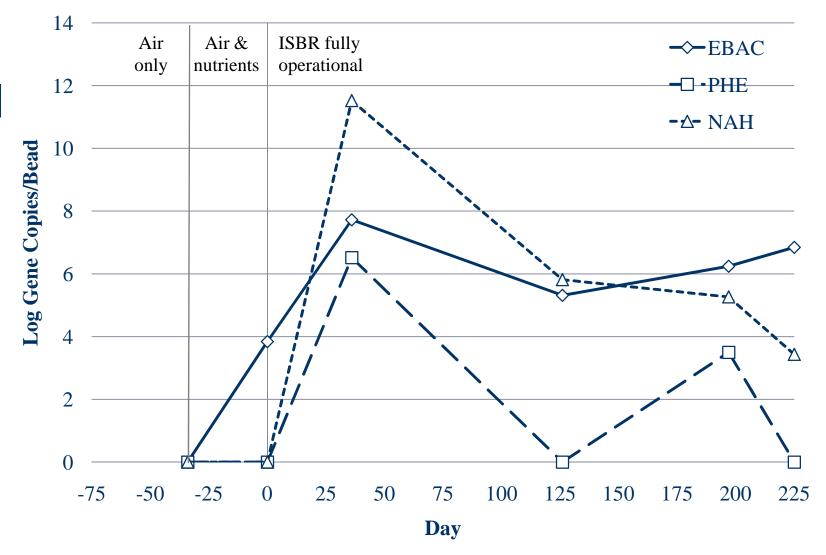
Comparison of NAH DNA and mRNA in BR1



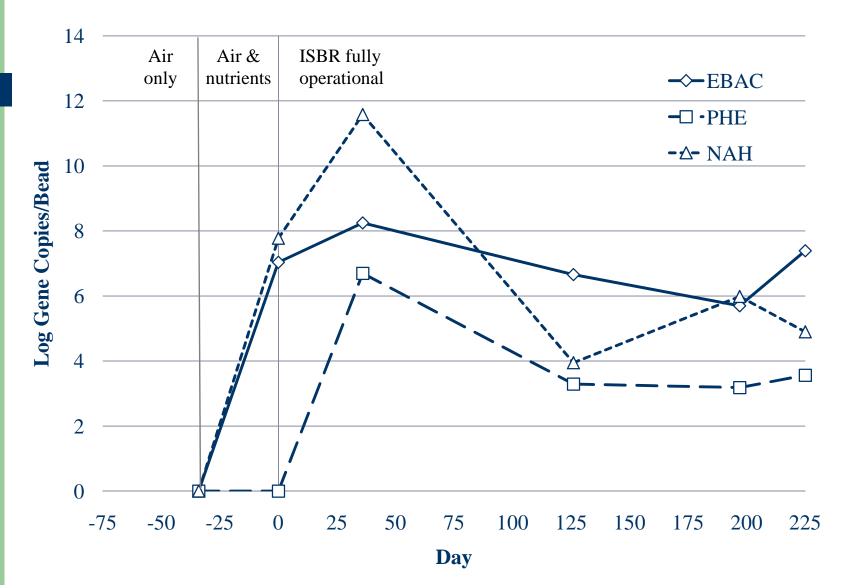
### **RT-qPCR results for BR1**



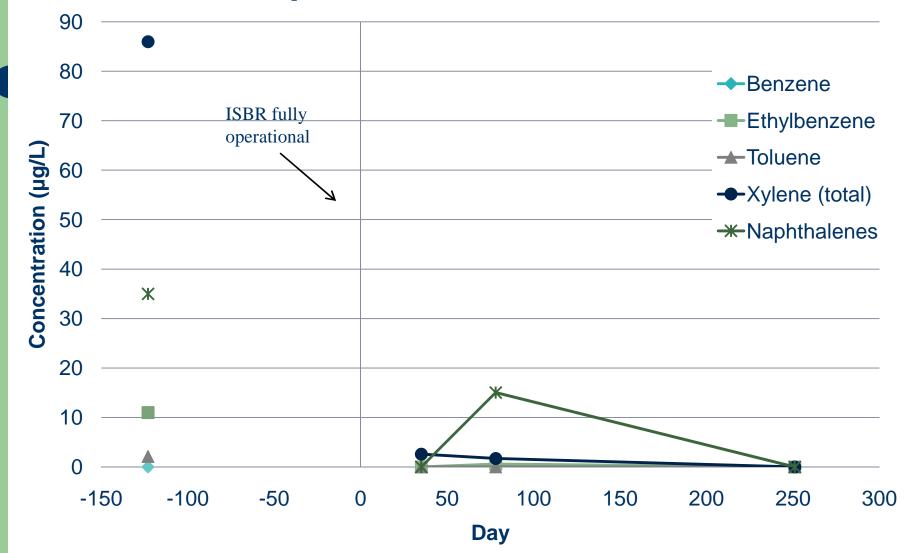
### RT-qPCR results for MW2, 1 ft downgradient from BR1



### RT-qPCR results for MW4, 2 ft downgradient from BR1



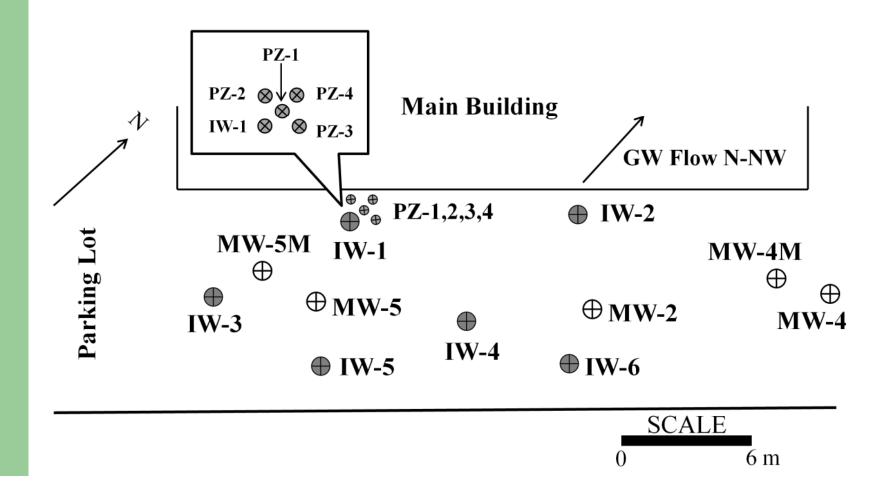
#### Concentrations of BTEX and Naphthalenes Over Time



### New Jersey site summary

- Increased concentration of degraders and increased expression of functional genes involved in degradation
- Microbial community responded to changes in treatment; effect is seen in both the bioreactor well and the monitoring wells
- All constituents of concern were either not detected or below groundwater quality criteria by the end of the study

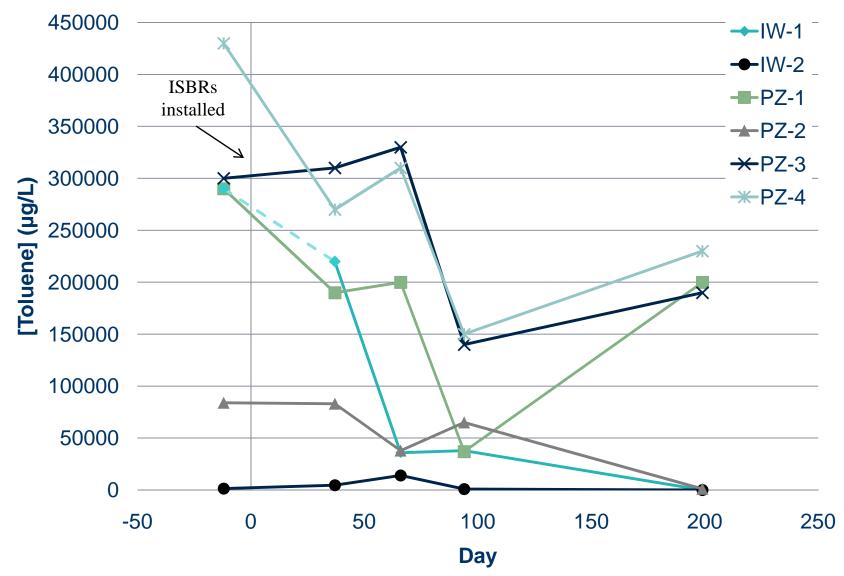
## Historic toluene release at a former industrial site



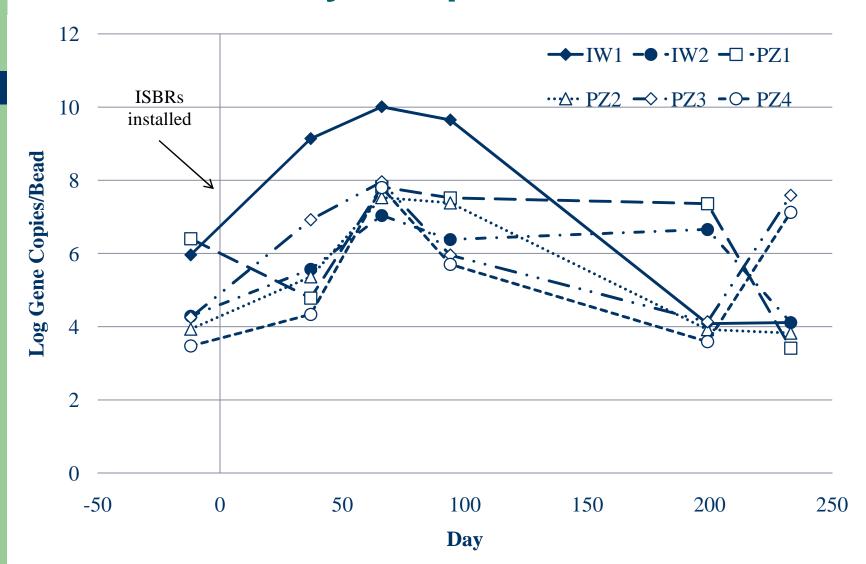
## Historic toluene release at a former industrial site

- Historic chemical releases, with up to 430 mg/L toluene
- Toluene concentrations above 250 mg/L shown to inhibit microbial activity
- Aquifer with high content of fine silt and clay
- Groundwater samples analyzed for contaminant concentrations
- Bio-traps deployed in each of the wells
  - RNA analysis and SIP

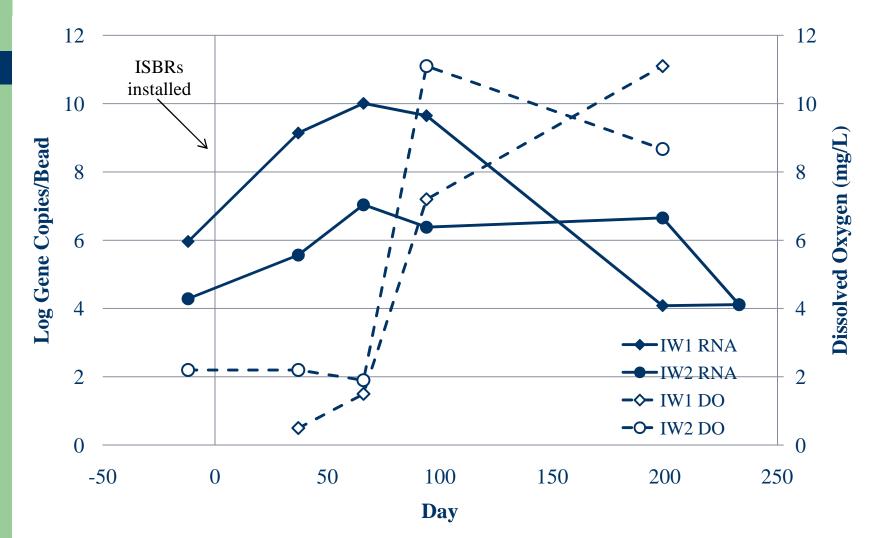
#### **Toluene Concentration**



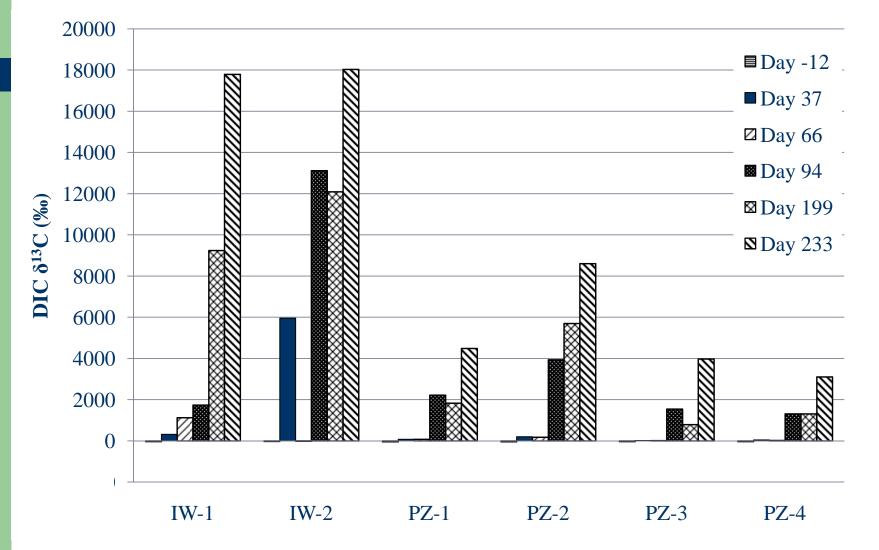
#### EBAC 16S rRNA genes detected by RT-qPCR



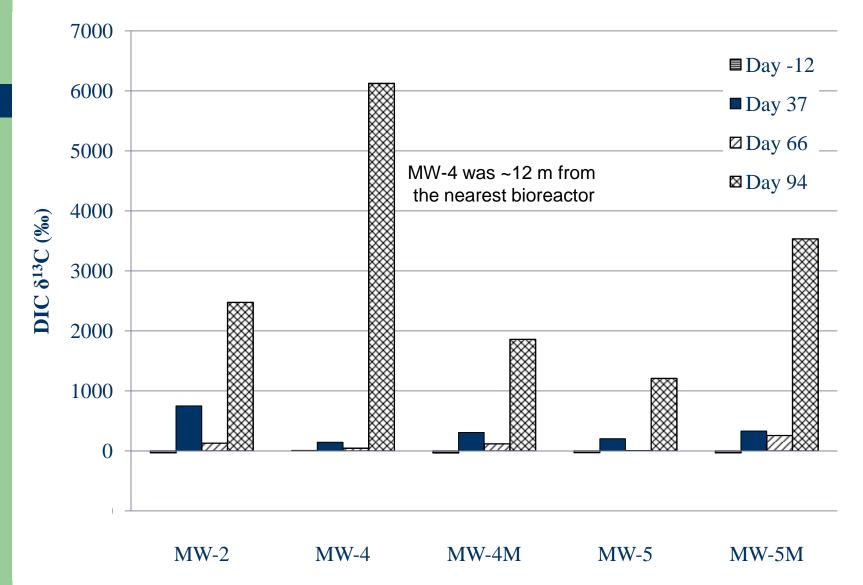
#### Total catabolic functional genes (NAH, PHE, TOD, RMO) detected by RT-qPCR and [DO]



#### DIC $\delta^{13}$ C values from IW-1, IW-2, and the PZ wells



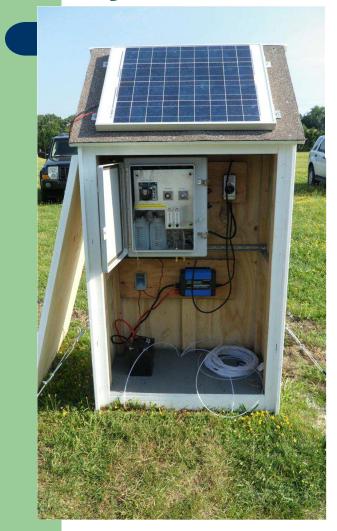
#### DIC $\delta^{13}$ C values from the MW wells



## Site summary

- By Day 199, toluene concentrations in all bioreactor wells met NJDEP groundwater standards
- Biodegradation was stimulated despite initial inhibitory toluene concentrations
- Toluene concentrations decreased between 31-47% in PZ-1, 3, & 4 and by 99% in PZ-2
- SIP data provided direct evidence of increased toluene mineralization in the bioreactor wells, adjacent PZ wells, and MW wells located throughout the site

## ISBR now being commercialized by



Engineering & Land Planning Associates, Inc.

## Acknowledgements

## This work would not have been possible without support from the following:





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## **Questions?**

